



Original Article

In-vitro Anticancer Activity of Ethyl Acetate Extract of *Aerva lanata* against MCF-7 Cell Line

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The aim of the present study is to evaluate the effect of in vitro anticancer activity of the ethyl acetate extract of *Aerva lanata* against MCF-7 human breast cancer cell line, using MTT assay showed a percentage of cell viability of 52% at 25 µg/ml which decrease with increase in concentration of extract. Anticancer activity of ethyl acetate extract of *Aerva lanata* on MCF-7 human breast cancer cell line showed potent cytotoxic activity. The inhibition percentage with regard to cytotoxicity was found to be 50% at 75µg/ml, which was comparable to the control Cyclophosphamide that showed a cytotoxicity of 49%. Therefore the minimum effective concentration of ethyl acetate extract of *Aerva lanata* was toxic to MCF-7 cells (IC₅₀) was recorded at a concentration of 45µg/ml of the ethyl acetate extract of *Aerva lanata*.

Key words: *Aerva lanata*, MCF-7 cell line, Ethyl acetate, MTT assay, cytotoxicity, Anticancer activity

1. INTRODUCTION

Aerva lanata Linn. belongs to the family Amaranthaceae, is a herbaceous perennial weed. The medicinal importance of *A. lanata* was used to treat diuretic, antiparasitic, anti helminthic¹, antidiabetic, expectorant and hepatoprotective², antimicrobial and cytotoxicity³, urolithiasis and anti-inflammatory activity⁴, nephroprotective⁵, anti-hyperglycemic⁶, Canthin-6-one and beta carboline alkaloids were isolated from *A.lanata* leaves⁷. The leaves of *A. lanata* are used as sap for eye complaints, an infusion is given to

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cure diarrhoea and kidney stone and the root is used in the snake bite.

Plant derived agents are being used for the treatment of cancer. Several anticancer agents from plants include, taxol, vinblastine, vincristine, the camptothecin derivatives, topotecan and irinotecan, and etoposide derived from epipodo phyllotoxin are in clinical use all over the world. Numerous cancer research studies have been conducted using traditional medicinal plants in an effort to discover new therapeutic agents that lack the toxic side effects associated with current chemotherapeutic agents. Scutellaria baicalensis was used as a component of PCSPES, an herbal mixture that showed efficacy in laboratory trials for prostate cancer, small-cell lung cancer and acute myeloid leukemia⁸⁻¹³.

The in vitro antitumor potential of the total alkaloid extract from *Tabebuia rosea* leaves against the human leukemic cells (MOLT-4) and the extract showed cytotoxic activity in a dose and time dependent manner¹⁴. The different extracts of hexane, chloroform, ethyl acetate and Crude methanol extract fractions of *Debregeasia salicifolia* stem for anticancer activity against MCF-7 cancer cell line and revealed that Mcf-7 showed minimum inhibition of 25.31% at the concentration of 10µg/ml and maximum inhibition of 99% was observed at the concentration of 500µg/ml¹⁵.

Deepa Philip, et al., (2011), have investigated the in vitro anticancer activity of *Sansevieria roxburghiana* against HepG2 liver cell and compared with normal 3T3 cells and showed a percentage of cell viability of 92.2% at 125µg/ml which decreased with increase in concentration¹⁶.

In vitro anticancer activity of *Rubia cordifolia* against HeLa and Hep2 cell lines and exhibited a significant cytotoxic activity in human cervical cancer cell line when compared to human larynx carcinoma¹⁷. The ethanolic extract of *Argemone mexicana*, *Polyalthia longifolia*, *Terminalia bellarica* and *Terminalia chebula* were evaluated for anticancer activity against HeLa-B75, Hep 3B and PN-15 cell lines and revealed that *P. longifolia* was found to be more potent against the HeLa cell lines¹⁸.

2. MATERIALS AND METHODS

Reagents

MEM was purchased from Hi Media Laboratories Fetal bovine serum (FBS) was purchased from Cistron laboratories Trypsin, methyl thiazolyl diphenyl- tetrazolium bromide (MTT), and Dimethyl sulfoxide (DMSO) were purchased from (Sisco research laboratory chemicals Mumbai). All of other chemicals and reagents were obtained from Sigma Aldrich Mumbai.

Media and Cell lines

Mcf-7 cell lines were obtained from National centre for cell sciences Pune (NCCS). The cells were maintained in Minimal Essential Media supplemented with 10% FBS, penicillin (100 µg/ml), and streptomycin (100 µg/ml) in a humidified atmosphere of 50 µg/ml CO₂ at 37 °C.

Collection of Plant material

Aerva lanata (includes leaf, stem and root), were collected from *Chinnapaliyampattu, Thiruvannamalai district*, the voucher specimen were kept in the Department of Zoology, Chennai, Tamil Nadu, India and used for this study.

Preparation of extract

The 25 g of dried powder of *A. lanata* was mixed with 100ml of ethanol solvent and kept in rotary shaker at 100 rpm overnight and filtered with whatmann no.1 filter paper and concentrated to dryness at 40⁰c. Until further use, different concentrations of the ethyl acetate extracts (25µg, 50µg, 75µg) were prepared in 5% Di-Methyl sulfoxide (DMSO) for determining cytotoxicity.

Cell viability assay on *Mcf-7* cells

The Cytotoxicity of samples on *Mcf-7* was determined by the MTT assay. Cells (1 × 10⁵/well) were plated in 100 µl of medium/well in 96-well plates (Costar Corning, Rochester, NY). After 48 hours incubation the cell reaches the confluence. Then, cells were incubated in the presence of various concentrations of the samples in 0.1% DMSO for 48h at 37°C. After removal of the sample solution and washing with phosphate-buffered saline (pH7.4), 20µl/well (5mg/ml) of 0.5% 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-tetrazolium bromide cells (MTT) phosphate- buffered saline solution was added. After 4h incubation, 0.04M HCl/isopropanol was added. Viable cells were determined by the absorbance at 450 nm. Measurements were performed and the concentration required for a 50% inhibition of viability was determined graphically. The absorbance at 450 nm was measured with a UV- Spectrophotometer using wells without sample containing cells as blanks. The effect of the samples on the proliferation of *Mcf-7* cells was expressed as the % cell viability, using the following formula:

% cell viability = A₄₅₀ of treated cells / A₄₅₀ of control cells × 100%.....(1)

The anticancer activity of ethyl acetate extract of *Aerva lanata* was performed on *Mcf-7* cell lines obtained from NCCLS Pune, India. The cell viability was measured using MTT assay as described above. Controls were maintained throughout the experiment. The assay was performed in triplicates for each of the extracts. The mean of the cell viability values was compared to the control to determine the effect of the extract. Cells and % viability was plotted against concentration of the plant extract. The maximum concentration of the plant extract that was toxic to *Mcf-7* was recorded as the effective drug concentration.

3. RESULTS AND DISCUSSION

The nontoxic dose of the ethyl acetate extract of *Aerva lanata* on *Mcf-7* showed that the percentage with regard to viability of cells was found to be 52% at a concentration of 25µg/ml which decreased with increase in concentration (Table 1). The extract showed a potent cytotoxic activity against *Mcf-7* cancer cell line (Table 1). Cyclophosphamide

served as positive control and 50% cancer inhibition was observed (fig2). The concentration of ethyl acetate extract of *Aerva lanata* at 75µg/ml showed an inhibition of 49% compared to that of positive control. Ethyl acetate extract of *Aerva lanata* at 25µg/ml, 50 µg/ml, 75 µg/ml showed cytotoxic activity of 47%, 48% and 59% respectively. Morphological changes of drug treated cells were examined using an inverted microscope and compared with the cells serving as control (Fig. 2). These observations may be due to the presence of active biological compounds. Therefore the minimum effective concentration of ethyl acetate extract of *Aerva lanata* was toxic to 50% *Mcf-7 breast* cancer cells was recorded (IC50) at a concentration of 75µg/ml of the plant extract.

Table 1: Cell viability assay on Mcf-7 cell line

| Concentrations(µg/ml) | % of Viability | % of Toxicity |
|-----------------------|----------------|---------------|
| Control | 100 | 100 |
| 25µg | 52.48 | 47.51 |
| 50µg | 51.624 | 48.37 |
| 75µg | 50.45 | 49.54 |
| Positive control | 50.5 | 49.5 |

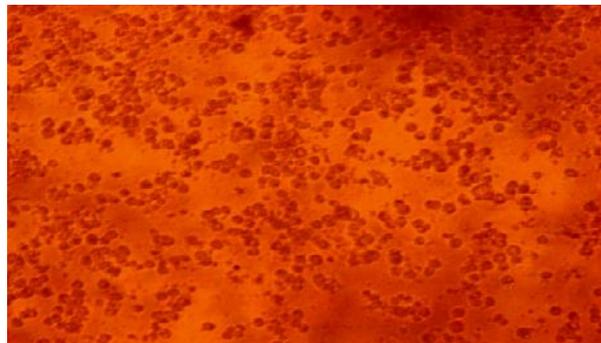


Fig 1: Positive Control

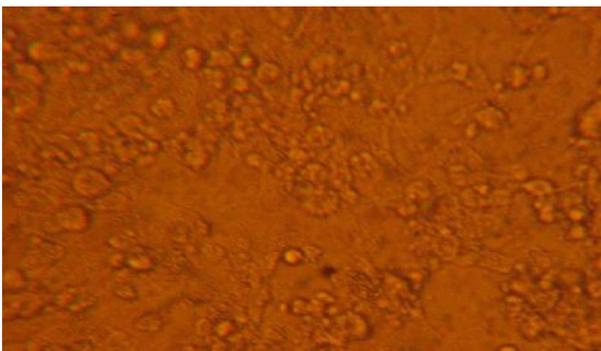


Fig 2: Ethyl acetate extract of Mcf-7 cell lines at 75µg

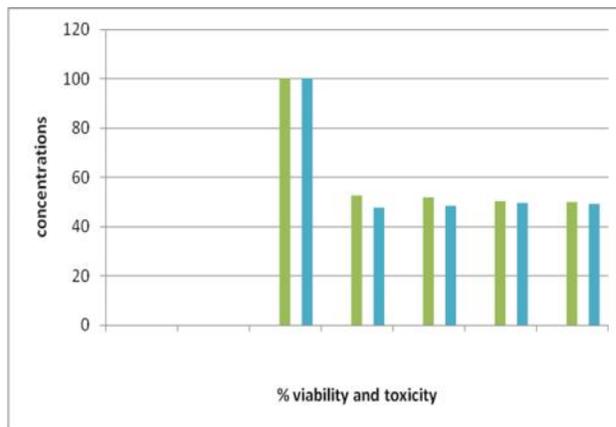


Fig 3: Percentage viability of Mcf-7 cell line

4. CONCLUSION

The results of this study support the efficacy of *Aerva lanata* as an anticancer agent for *Mcf-7 breast* cancer cell line. From the present study it has been revealed that ethyl acetate extract of *Aerva lanata* shows 50% anticancer activity in *Mcf-7* cancer cell line at the concentration of 75µg/ml. It acts a potential adjuvant treatment to current chemotherapeutic agents and can be used in the treatment of *Mcf-7* and a further research has to be done. From this it is said that due to the presence of *phyto component s*¹⁹, it shows 50% activity. In future the components present on *Aerva lanata* may act as a drug, further in-vivo studies should be carried out. Considerable works have been done on the medicinal plants to treat cancer, and some plant products have been marketed as anticancer drugs. These plants may promote host resistance against infection by re-stabilizing body equilibrium and conditioning the body tissues.

5. REFERENCES

1. Anantha D, Israiel kumar T, Santhosh kumar M, Manohar reddy A, Mukharjee MSV and Lakshmana rao A. Invitro antihelminthic activity of aqueous and alcoholic extract of Aerva lanata seeds and leaves. J.Pharm. Sci and Res. 2010; 2(5): 317 – 321.
2. Kirtikar KR and Basu MD, Indian Medicinal Plants. Second edition. Periodical express book agency. New Delhi. 1991, 3: 1926.
3. Dulay C. Antimicrobial activity and cytotoxicity of Aerva lanata. Fitoterapia, 2002; 73: 93 – 94.
4. Vetrichelvan T, Jagadeesan M, Senthil Palaniappan S, Murali MP and Sasikumar K. Diuretic and anti-inflammatory activities of *Aerva lanata* in rats. Indian J Pharm. 2000; 62: 300 – 302.
5. Manokaran S, Jaswanth A, Sengotuvelu S, Nandhakumar J, Duraisami R, Karthikeyan D and Mallegaswari R. Hepatoprotective activity of Aerva lanata against paracetamol induced hepatotoxicity in rats. Research J Pharm and Tech 2008; 1(4); 398 – 400.

6. Shirwaikar A, Issac D, Malini S. effect of *Aerva lanata* on Cisplatin and gentamicin models of acute renal failure. *J.Ethnopharmacol.* 2004; 90: 81 – 86.
7. Zapesochay G. Canthin-6-one and beta carboline alkaloids from *Aerva lanata*. *Planta medica.* 1992; 58: 192 -196.
8. Chung VQ, Tattersall M, Cheung HT. Interactions of a herbal combination that inhibits growth of prostate cancer cells. *Cancer Chemother Pharmacol.* 2004; 53:384-390.
9. Cordell GA. PC-SPES: a brief overview. *Integr Cancer Ther.* 2002; 1:271-286.
10. Hsieh TC, Lu X, Chea J, Wu JM. Prevention and management of prostate cancer using PC-SPES: a scientific perspective. *J Nutr* 2002; 132:3513S-3517S.
11. Ikezoe T, Chen S, Saito T, Asou H, Kyo T, Tanosaki S, Heber D, Taguchi H, Koeffler HP: PC-SPES decreases proliferation and induces differentiation and apoptosis of human acute myeloid leukemia cells. *Int J Oncol.* 2003; 23:1203-1211.
12. Meyer JP, Gillatt DA. PC-SPES: a herbal therapy for the treatment of hormone refractory prostate cancer. *Prostate Cancer Prostatic Dis.* 2002; 5:13-15.
13. Oh WK, Kantoff PW, Weinberg V, Jones G, Rini BI, Derynck MK, Bok R, Smith MR, Bublely GJ, Rosen RT, DiPaola RS, Small EJ: Prospective, multicenter, randomized phase II trial of the herbal supplement, PC-SPES, and diethylstilbestrol in patients with androgen-independent prostate cancer. *J Clin Oncol.* 2004; 22:3705-3712.
14. Deepa Philip A, Kaleena P and Valivittan K. Invitro Cytotoxicity and Anticancer Activity Of *Sansevieria roxburghiana*. *International Journal of Current Pharmaceutical Research.* 2011; 3(3): 71-73.
15. Mani Sathiyaa, Krishnaswamy Muthuchelian.. Antitumor Potential of Total Alkaloid Extract from *Tabebuia rosea*(Bertol.) DC. Leaves on MOLT-4 Cells In Vitro. *Nature and Science* 2010;8(9): 77-85.
16. Parag R. Patel, Akhil A. Nagar1, Rikin C. Patel, Dhara K. Rathod1, Vishal R. Patel. Invitro Anticancer Activity of *Rubia cordifolia* Against Hela and Hep2 Cell Lines. *International Journal Of Pharmacy And Pharmaceutical Sciences.* 2011; 3(2).70-71.
17. Rajesh N, Gacche, Rafik U Shaikh, Mahesh M Pund. Invitro evaluation of anticancer and antimicrobial activity of selected medicinal plants from Ayurveda *Asian journal of Traditional medicines.* 2011; 6(3): 117-133.
18. Sobia Nisa, Yamin Bibi, Abdul Waheed, Muhammad Zia, Sadia Sarwar, Sabbir Ahmed and M. Fayyaz Chaudhary. Evaluation of anticancer activity of *Debregeasia salicifolia* extract against estrogen receptor positive cell line. *African Journal of Biotechnology.* 2011;10(6), 990-995.
19. Krishnamoorthi R: Phytochemical Analysis and Antioxidant Property of *Aerva Lanata*. *Int J Pharmacognosy* 2015; 2(8): 426-29

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